Molecular Simulations for Anti-amyloidogenic Effect of Flavonoid Myricetin Exerted against Alzheimer’s β-Amyloid Fibrils Formation

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Comparative molecular simulations were performed to establish molecular interaction and inhibitory effect of flavonoid myricetin on formation of amyloid fibrils. For computational comparison, the conformational stability of myricetin with amyloid β-peptide (Aβ) and β-amyloid fibrils (fAβ) were traced with multiple molecular dynamics simulations (MD) using the CHARMM program from Monte Carlo docked structures. Simulations showed that the inhibition by myricetin involves binding of the flavonoid to fAβ rather than Aβ. Even in MD simulations over 5 ns at 300 K, myricetin/fAβ complex remained stable in compact conformation for multiple trajectories. In contrast, myricetin/Aβ complex mostly turned into the dissociated conformation during the MD simulations at 300 K. These multiple MD simulations provide a theoretical basis for the higher inhibitory effect of myricetin on fibrillogenesis of fAβ relative to Aβ. Significant binding between myricetin and fAβ observed from the computational simulations clearly reflects the previous experimental results in which only fAβ had bound to the myricetin molecules.

Key Words: Amyloid fibril, Conformational analysis, Flavonoid, Molecular dynamics simulations, Myricetin

Introduction

Amyloidosis is a clinical condition marked by the deposition of fibrous amyloids in various organs and tissues of the body.1 The protein fibril formation may be causative for various fatal human disorders including Alzheimer’s disease (AD).2 Common structural feature of amyloid fibrils has been characterized by critical cross β-sheet core structure that perpetuates along the fibril long axis.3 Fibrillogenesis of the monomeric amyloid β-peptide (Aβ) found in AD generally occurs via nucleation-dependent polymerization.4 If supersaturated Aβ solution forms a nucleus, the growth of fibril is accelerated to evolve into the β-amyloid fibrils (fAβ).5 The structure-toxicity relationship study performed by Riek and co-workers indicates that fAβ contain an intrinsic toxicity that correlates with their morphologies as well as quantities.6 They showed that the neurotoxicity was generally proportional to the size of fAβ. Therefore, it would be crucial to regulate the fAβ growth by chemical additives to develop potent anti-Alzheimer’s disease drugs.

Recent studies for the fAβ fibrillation revealed that some flavonoid compounds are able to inhibit the fibril growth and destabilize the preformed fAβ.7 Ono et al. found that myricetin exhibited its anti-amyloidogenic effect via preferential binding to the fAβ rather than the Aβ monomers.8 In order to develop anti-amyloidogenic drugs, it is mandatory for structural biologists to acquire a detailed understanding of molecular interactions between flavonoids and fAβ.

In this report, comparative molecular simulations were performed for both myricetin/Aβ and myricetin/fAβ in order to explain the experimentally observed inhibitory effect of myricetin for the fibril growth of Aβ. To reproduce real interactions between myricetin and amyloid peptide, Monte Carlo (MC) docking and molecular dynamics (MD) simulations were carried out for the molecular models of myricetin/Aβ and myricetin/fAβ. Significant binding between myricetin and fAβ observed from the computational simulations clearly reflects the previous experimental results9 in which only fAβ bind to the myricetin molecules. From the simulation of fibrillar fAβ with myricetin, it has been found that the myricetin interactions with the outermost part of fibrils are responsible for the experimentally observed inhibitory effect on the fibrillogenesis of fAβ.

Results and Discussion

Differences in docked conformations and energetics of the myricetin with Aβ and fAβ. The interaction energy profiles of each myricetin complex with Aβ and fAβ during MC docking simulations are shown in Figure 1. The intermolecular energy between myricetin and Aβ was −18.83 kcal/mol and those of myricetin and fAβ was −21.41 kcal/mol during last 6 × 10⁵ MC trials. This means that the myricetin molecule forms a more stable complex with fAβ than the monomeric Aβ. This result seems to be consistent
with the experimental observation that the flavonoid myricetin exerted its binding ability only to fAβ rather than Aβ. Interaction energy profiles of the myricetin/Aβ and the myricetin/fAβ are shown in Figure 2. Myricetin was docked onto a region between α-helix and C-termini of the Aβ. There was no special binding site on the Aβ and the myricetin just clung to the surface of the peptide (Fig. 2). Contrary to the case of myricetin/Aβ, the myricetin formed a stable docked complex with the fAβ. The U-shaped conformation of fAβ provided suitable binding site for a flat shaped myricetin molecule. The myricetin was fully stuck into the hydrophobic binding patch of the fAβ. These two conformations were further investigated by MD simulations.

**Complex stability of myricetin/Aβ and myricetin/fAβ during multiple MD simulations.** Multiple MD simulations were performed from the lowest energy conformation of MC simulations. These MD simulations for each myricetin complex evidenced more stable conformation of myricetin/fAβ compared to myricetin/Aβ. Figure 3 shows center of mass distance between myricetin and Aβ or fAβ during 5-ns of MD simulations for six trajectories. Four trajectories showed gradual dissociation of the myricetin from Aβ with some variations (Figure 3a-d). Stable docked conformations were only maintained for just two trajectories with showing high-fluctuations (Figure 3e, f). Therefore, we can conclude that the interaction between myricetin and Aβ may be weak and unsuitable for keeping the complex stability during MD simulations. Compared with the myricetin/Aβ complex, the myricetin showed a more stable structure with the fAβ during multiple MD simulations. Final dissociation of myricetin from fAβ were observed for only two trajectories (Figure 4c, d). Others were tightly associated with each other during MD simulations, in which averaged center of mass distances were 0.83, 0.70, 0.53, and 0.52 nm, respectively (Figure 4a, b, e, f).

The differences in the conformational stability of the myricetin/Aβ and the myricetin/fAβ were also supported by minimum distances between the myricetin and Aβ or fAβ (Figure 5). After 5-ns of MD simulations, trajectory-averaged minimum distance between myricetin and Aβ was about 3 nm, while the distance between myricetin and fAβ was below 2 nm. This means that the atomic interactions between myricetin and fAβ were more stable than those of myricetin and Aβ. Figure 6 is a solvent-accessible surface area change upon formation of complexes between myri-
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At early MD phase (0-1 ns), the surface area change from binding of myricetin to fAβ was −5.85 nm². This indicates that the surface area was decreased by approximately 5.85 nm² due to the binding of myricetin and fAβ. The value was increased to −3.70 nm² during late MD simulation phase (4-5 ns). For the myricetin/Aβ, the surface area change was −4.40 nm² during early MD phase and the value was increased to −1.33 nm² during late MD phase. The surface area change of myricetin/fAβ was much lower than that of myricetin/Aβ at any MD time phase. These results suggest tighter binding of myricetin to the fAβ in comparison to Aβ. The ratio of surface area changes during late to early MD phase was 0.63 for the myricetin/fAβ and 0.30 for the myricetin/Aβ. This means that the surface area decrease of myricetin/fAβ was two-fold higher than that of myricetin/Aβ during the time scale of MD simulations. Therefore, binding stability of myricetin to the fAβ is more tolerant to dynamic fluctuation of the molecular complex during the MD simulation time. The strong binding ability of myricetin to fAβ in comparison to Aβ could provide a molecular understanding for experimentally observed inhibitory effect of myricetin on amyloid fibrilligenesis.

Figure 4. Center of mass distances between myricetin and fibrous form of amyloid during multiple 5ns-MD simulations.

Figure 5. Averaged minimum distance between myricetin and helical (red line) or fibrous (blue line) form of amyloid peptide during multiple MD simulations.

Figure 6. Averaged surface area change upon complexation of myricetin with helical (red line) or fibrous (blue line) form of amyloid peptide during multiple MD simulations.

Figure 7. Destabilization of fAβ fibrils with myricetin (a) and stable fAβ fibrils (b) during 10ns MD simulations.
Destabilization of fAβ by myricetin during the MD simulations. Myricetin can destabilize existing fibrils as well as have inhibitory effect on formation of emerging fibrils. Figure 7 is the time course of the fibril destabilization by myricetin during 10-ns of MD simulations. The fibrils were modeled as five sets of fAβ monomers. After 280 ps, the myricetin made an interaction with the outermost fAβ sheaf. After 550 ps, the myricetin had bound to C-termini of the fAβ sheaf via hydrophobic interaction of aromatic rings. The binding interaction was intensified into the core cavity of the outermost fAβ sheaf after 1.85 ns of MD simulation. In that time, about half of the domain of fAβ sheaf was segregated from remaining fAβ fibrils. The level of interaction between fibrils and fAβ part was minimized by the bound myricetin. After 2.69 ns, the myricetin moved out of N-termini of fAβ sheaf and showed relatively decreased interaction with the fAβ sheaf. Finally, the myricetin moved to the next part of fAβ sheaf to destabilize remaining part of fAβ fibrils after 8.11 ns. From the MD simulation snapshots in the Figure 7, it is evidenced that the myricetin exert an anti-amyloidogenic effect by destabilizing the fAβ fibrils as well as inhibiting the formation of fibrils from monomeric peptides.

For the quantitative comparison between with and without myricetin conditions, Cα atomic distance was analyzed for each fAβ sheaf in the fibrils. Figure 8 is time course of interatomic distances between fAβ sheaf 1-2, 2-3, 3-4, and 4-5 for centric Cα atom of each Gly13 residue. During the MD simulations, it was found the interatomic distances of outer fAβ sheaf 1-2 or 4-5 were dramatically increased by myricetin (Figure 8a, d). However the distances for the inner fAβ sheaf were not changed during the MD simulations for both with and without myricetin conditions (Figure 8b, c). It seems that myricetin moleculer exert anti-amyloidogenic effect for outer part of fAβ fibrils by perturbing these inter-sheaf vdw interactions. Figure 9 is intermolecular electrostatic and vdw energy between outermost fAβ sheaf 1 and 2.

During MD simulations, the intermolecular electrostatic energies were controversial to each fAβ fibril with and without myricetin (Figure 9a). However, intermolecular vdw energies were discriminated between the fAβ fibrils (Figure 9b). The vdw energy of fAβ fibrils was lowered than the case of fAβ fibrils with myricetin system during the simulation times. Intermolecular vdw energy was $-91.2$ kcal/mol for the fAβ fibrils without myricetin and $-71.3$ kcal/mol for the fAβ fibrils with myricetin, respectively. That means myricetin molecule destabilize the intermolecular vdw interaction between outer part of fAβ sheaf rather than interior part of fAβ fibrils.

Methods of Computation

Construction of the molecular models and protocol of MC docking simulations. The starting configuration of each Aβ and fAβ for MC simulations was obtained from the Protein Data Bank (PDB). The PDB ID’s for Aβ and fAβ were 1zoq and 2beg respectively. To obtain representative conformations of the 3D coordinate ensemble, the Discovery Studio/Builder module (version 1.7, Accelrys Software Inc.) was used as a molecular editor. The atomic charges and atomic partial charges of the myricetin were obtained by utilizing ab initio calculations with 6-311G basis set at Hartree-Fork (HF) level. All simulations were performed using a general molecular modeling program, CHARMM (version 30b1), with a parm22 all-atom force field. The MC docking simulations were performed using the “MC” module.
of CHARMM. The short-range non-bonding interactions were truncated with a 13-Å cutoff. An implicit solvent water model was used with a distance-dependent dielectric constant. The docking process was assumed to be a 1:1 interaction between each amyloid peptide and myricetin during the MC runs. The initial configuration of each amyloid and myricetin molecule was positioned arbitrarily within a neighboring distance. Trials to a new configuration were accomplished by changing each move set of a guest molecule. The MC move set for flexible docking was composed of rigid translations, rigid rotations, and rotations of freely rotatable dihedral angles of the both myricetin and amyloids. A single step consists of picking a random conformer, making a random move, minimizing the energy of a new conformer, and then checking the energy with a Metropolis criterion. The MC-accepted structures were saved every 4000 steps for 8,000,000 trials. Multiple 10-MC trajectories were generated with different random number seed to convergence of the simulation. These MC processes produced convergent docked structures for each monomeric amyloid with myricetin.

Multiple molecular dynamics simulations of the each myricetin/Aβ and myricetin/fAβ complexes. The starting configurations of each myricetin/β and myricetin/fAβ complex for the MD simulations in explicit water were taken from the MC-docked conformations with the lowest-energy value (Figure 2). The geometries of these molecular models were fully optimized before MD runs. A TIP3P three-site rigid water model was used to solvate the complexes. Water molecules were removed if they were closer than 2.8 Å to any heavy atoms of the complexes. In summary, each system was constructed using periodic boundary conditions with a cubic box of dimensions 50 Å × 50 Å × 50 Å, consisting of myricetin/β and 4,320 water, or myricetin/fAβ complex and 4,293 water molecules. The system was minimized by 2,000 steps of conjugate gradient, followed by Adopted Basis Newton-Raphson until the root-mean-square gradient was less than 0.001 kcal/mol. The MD simulations were performed using the CHARMM 30b1 program in the isothermal-isobaric ensemble at 300 K (P = 1 bar, T = 300 K). The particle mesh Ewald summation method was used to treat the long-range electrostatic interactions. The bond lengths of all molecules were constrained with the SHAKE algorithm. The time step was 2.0 fs, and the non-bonded pair list was updated every 25 steps. The short-range non-bonded interactions were truncated with a 13-Å cutoff. The temperature and pressure of the system were regulated using the Langevin piston method in conjunction with Hoover’s thermostat. The system was gradually heated to each targeted temperature for 50 ps and equilibrated at this temperature. The production MD trajectory with one snapshot per 2 ps was collected for 5,000 ps. Total six-independent MD simulations were performed with different random number seed to cover wide conformational sampling. For the simulation of whole fibrils with myricetin, molecular model was constructed with the cubic box of dimensions 70 Å × 70 Å × 70 Å, consisting of four myricetins, five-fAβ fibril, and 10,216 water molecules. The MD simulations were carried out over the course of 10-ns to observe destabilization of fAβ fibril by myricetin.

Conclusions

Comparison of the dynamics of myricetin/Aβ and myricetin/β complex during molecular simulations demonstrate that myricetin disturb upon Alzheimer’s β-amyloid fibrillogenesis by preferential binding into the hydrophobic patch of fAβ. Multiple MD simulations of myricetin/Aβ and myricetin/fAβ provided the important theoretical insight into the experimentally determined anti-amyloidogenic effect of flavonoid compound, suggesting that myricetin could bind to fibrous form of amyloid peptides, thereby decreasing favorable interaction between monomeric β-sheets of amyloid fibrils. Based on the present study, we conclude that computational methodology can make great contribution toward anti-amyloidogenic drug design using natural flavonoid compounds.

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