Antioxidant and Neuronal Cell Protective Effects of Columbia Arabica Coffee with Different Roasting Conditions

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ABSTRACT: In vitro antioxidant activities and neuronal cell protective effects of ethanol extract from roasted coffee beans were investigated. Colombia arabica coffee (Coffea arabica) green beans were roasted to give medium (230 oC, 10 min), city (230 oC, 12 min) and french (230 oC, 15 min) coffee beans. Total phenolics in raw green beans, medium, city and french-roasted beans were 8.81±0.05, 9.77±0.03, 9.92±0.04 and 7.76±0.01 mg of GAE/g, respectively. The content of 5-O-caffeoylquinic acid, the predominant phenolic, was detected higher in medium-roasted beans than others. In addition, we found that extracts from medium-roasted beans particularly showed the highest in vitro antioxidant activity on ABTS radical scavenging activity and FRAP assays. To determine cell viability using the MTT assay, extracts from medium-roasted beans showed higher protection against H2O2-induced neurotoxicity than others. Lactate dehydrogenase (LDH) leakage was also inhibited by the extracts due to prevention of lipid peroxidation using the malondialdehyde (MDA) assay from mouse whole brain homogenates. These data suggest that the medium-roasting condition to making tasty coffee from Columbia arabica green beans may be more helpful to human health by providing the most physiological phenolics, including 5-O-caffeoylquinic acids.

Keywords: Coffea arabica, coffee, neurotoxicity, roasting condition, 5-O-caffeoylquinic acid

INTRODUCTION

Reactive oxygen species (ROS) such as hydrogen peroxide (H2O2), singlet oxygen, superoxide anion radical (O₂−) and hydroxyl radical (OH−) are generated from the autoxidation of lipids, as well as reactive nitrogen species (RNS) (1). Formations of these excess ROS and RNS by UV irradiation, smoking and drug metabolism are likely to damage several cellular components such as lipids, proteins, nucleic acids and DNA through oxidation or nitration processes (2). In addition, these reactive oxygen species cause inflammation or lesions on various organs, and are associated with various degenerative diseases, including cancer, aging, arteriosclerosis and neurodegenerative diseases (3,4). Alzheimer’s disease (AD) is one of the most serious threats to human health in aged societies of developed countries. In particular, AD, one of the major neurodegenerative diseases, is characterized by loss of memory and cognition. After amyloid plaque formation, inflammation and oxidative stress further enhance the degeneration of neurons. Accumulation of intracellular H2O2 induces the peroxidation of membrane lipids and apoptotic cell death by activation of caspases (5,6). However, some phytochemicals from natural plant sources like fruits and vegetables may reduce the risk of AD because of their antioxidative properties diminishing oxidative insults (7).

Coffee has been one of the most popular beverages all over the world, and its consumption continues to increase due to its physiological effects as well as its pleasant taste and aroma. In addition, coffee is one of the most important food commodities both for producers, in tropical and subtropical areas with coffee as their main agricultural export product, and for manufacturers, which are mainly located in Europe and North America where coffee is roasted, mixed and packed. Due to its large diffusion and high market value, coffee is subject to adulteration throughout its production chain (8).

Generally, coffee has been reported to have beneficial effects on degenerative diseases including cancer (9), cardiovascular disorders (10), obesity and diabetes (11,12). Coffee contains phenolic acids including caffeic acid,
ferulic, and vanillic acid (13), and 5-O-caffeoylquinic acid as the predominant phenolic compound, measuring on average 100 mg per cup of coffee (14). Although roasted coffee has physiological activities because of their phytochemicals, little is known about the difference in physiological activities due to various roasting conditions for coffee beans. In addition, many complex physical and chemical changes take place during roasting from the obvious change in color from green to dark brown, to the major compositional changes such as decreases in protein, amino acids, arabinoxylan, reducing sugars, trigonelline, 5-O-caffeoylquinic acid, sucrose, and water, and the formation of melanoidins (15).

The objective of this experiment was to investigate the effects of coffee beans with various roasting conditions against oxidative stress-induced neurotoxicity.

**MATERIALS AND METHODS**

**Materials**
Folin-Ciocalteu’s phenol reagent, 2,2’-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS), potassium persulfate, 2,4,6-tripyridyl-S-triazine (TPTZ), trichloroacetic acid (TCA), thiobarbituric acid (TBA), vitamin C, α-tocopherol, catechin, 2-[4-(2-hydroxyethyl)piperazin-1-yl]ethanesulfonic acid (HEPES), sodium bicarbonate, penicillin, streptomycin, 5-O-caffeoylquinic acid, syringic acid, epigallocatechin gallate, ferrous sulfate (FeSO4), hydrogen peroxide (H2O2), dimethyl sulfoxide (DMSO), 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide (MTT) assay kit, lactate dehydrogenase (LDH) assay kit and all solvents used were of analytical grade and purchased from Sigma Chemical Co. (St. Louis, MO, USA). RPMI 1640 medium and fetal bovine serum were obtained from Gibco BRL (Grand Island, NY, USA).

**Sample preparation**
Colombian arabica coffee (Coffee arabica) beans were purchased from the local barista institute in Jinju of Korea, in December 2010. The sample was roasted for 10, 12 and 15 min at 230°C to give medium-, city- and Line Bank, Seoul, Korea) were propagated in RPMI 1640 medium containing 10% fetal bovine serum, 50 units/mL penicillin and 100 μg/mL streptomycin.

**ABTS radical scavenging activity**
2,2’-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid) (ABTS) was dissolved in water to make a concentration of 7 mmol/L. ABTS was produced by reacting the ABTS stock solution with 2.45 mM potassium persulfate (final concentration) and allowing the mixture to stand in the dark at room temperature for 12~16 hr before use. For the study of samples, the ABTS stock solution was diluted with phosphate-buffered saline (5 mM/L, pH 7.4) to an absorbance of 0.70 at 734 nm. After the addition of 980 μL of diluted ABTS to 20 μL of sample, the absorbance reading was taken 5 min after the initial mixing (16) and the percent ABTS scavenging activity is calculated as:

% ABTS scavenging activity = [(control absorbance − sample absorbance)/(control absorbance)] × 100

**Ferric reducing antioxidant power (FRAP)**
To measure total antioxidant activity, the FRAP assay, developed by Jeong et al. (16), was used. In short, 1.5 mL of working, pre-warmed 37°C FRAP reagent (10 volumes of 300 mM sodium acetate buffer (pH 3.6), 10 mM 2,4,6-tripyridyl-S-triazine in 40 mM HCl, and 20 mM FeCl3 · 6H2O) was mixed with 50 μL of test samples and standards. The mixture was vortexed and allowed to stand in the dark at room temperature for 12 hr. After dilution with phosphate-buffered saline (5 mM/L) and allowing the mixture to stand in the dark at room temperature for 12~16 hr before use. For the study of samples, the ABTS stock solution was diluted with phosphate-buffered saline (5 mM/L, pH 7.4) to an absorbance of 0.70 at 734 nm. After the addition of 980 μL of diluted ABTS to 20 μL of sample, the absorbance reading was taken 5 min after the initial mixing (16) and the percent ABTS scavenging activity is calculated as:

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**Malondialdehyde (MDA) assay using mouse whole brain homogenates**
The MDA assay was carried out to the method described by Chang et al (17). The brain of young adult male ICR mice were disected and homogenized in ice-cold Tris-HCl buffer (20 mM, pH 7.4) to produce a 1/10 homogenate. The homogenate centrifuged at 12,000xg for 15 minutes at 4°C. 1 mL aliquots of the supernatant were incubated with the test samples in the presence of 10 μM FeSO4 and 0.1 mM vitamin C at 37°C for 1 hr. The reaction was terminated by addition of 1.0 mL TCA (28%, w/v) and 1.5 mL TBA (1%, w/v) in succession, and then the solution was heated at 100°C. After 15 minutes, the color of the MDA-TBA complex was measured at 532 nm. (+)-Catechin, a well-known antioxidant, was used as a positive control. The inhibition ratio (%) was calculated as follows:

% inhibition = [(absorbance of control − absorbance of sample)/absorbance of control] × 100
**Determination of cell viability**

[3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazoliumbromide] (MTT) reduction assay was determined using the in vitro toxicology assay kit (TOX-1, Sigma Co.). Neuronal PC12 cells were plated at a density of $10^5$ cells/well in 96-well plates in 100 μL of RPMI. The cells were pre-incubated with various extracts obtained from roasted coffee for 48 hr before the cells were treated either with or without 200 μM H₂O₂ for 2 hr. The amount of MTT formazan product was determined using absorbance using a microplate reader (680, Bio-Rad, Tokyo, Japan) at a test wavelength of 570 nm and a reference wavelength of 690 nm (17).

Neuronal PC12 cells were precipitated by centrifugation at 250×g for 4 min at 4°C, 100 μL of the supernatants were transferred into new wells, and lactate dehydrogenase (LDH) was determined using the in vitro toxicology assay kit (TOX-7, Sigma Co.). Damage of the plasma membrane was evaluated by measuring the amount of the intracellular LDH enzyme released into the medium (18).

**Determination of total phenolics**

Total phenolics were determined by spectrophotometric analysis (16). The standard curve for total phenolics was made using 5-O-caffeoylquinic acid standard solution (0~100 mg/L). Total phenolics in roasted coffee were expressed as milligrams of gallic acid equivalents per gram (mg GAE/g) of sample.

**Quantification by HPLC**

Phenolics in extracts obtained from roasted coffee beans were measured at 280 nm by a photo diode array detector (Ultimate 3000 series, Dionex, Sunnyvale, CA, USA). Separation was achieved with a Shiseido C18 column (250 mm×4.6 mm ID×5 μm, Shiseido Co., Tokyo, Japan). The elution solvents were (A) 0.01 M potassium phosphate buffer adjusted to pH 3.0 by phosphoric acid and (B) methanol (99.9% concentrated). The solvent gradient elution program used was as follows: initial 90% (A), hold for 9.5 min; linear gradient to 68% (A), 3.5 min; linear gradient to 67% (A), 17 min; linear gradient to 20% (A), 1 min; linear gradient to 90% (A), 1 min, and hold for 10 min. The flow rate was 1.5 mL/min. Phenolics were identified by comparing their retention time (RT) values to UV spectra of known standards and quantified by peak areas from the chromatograms. All analyses were run in triplicate and mean values were calculated. Content of phenolic compounds was expressed in mg/g extract.

**Statistical analysis**

All data were expressed as mean±SD. Each experimental set was compared with one-way analysis of variance (ANOVA) and Duncan’s multiple-range test (p<0.05) using SAS program (SAS Institute Inc., Cary, NC, USA).

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**RESULTS AND DISCUSSION**

**ABTS radical scavenging activities and FRAP**

The reduction capability of ABTS induced by antioxidants was determined by the decrease in its absorbance at 734 nm. A positive ABTS test suggests that the extracts were free radical scavengers. The scavenging effect of extracts obtained from roasted coffee and vitamin C on ABTS radicals were compared. The extracts had significant ABTS radical scavenging effects with increasing concentrations in the range of 125~1,000 μg/mL. A 1,000 μg/mL concentration of the extracts (green, medium, city and french) exhibited 46.99±5.40%, 59.85±1.98%, 55.27±0.08% and 56.5±1.22% inhibitions, respectively (Fig. 1A). However, when compared with that of vitamin C as a positive control, the scavenging effect of the extracts was lower. These results showed that extracts obtained from roasted coffee had ABTS radical scavenging activities, though the activities of the tested samples were lower than that of vitamin C.

Antioxidants can be referred to as reductants, which inactivate oxidants, and are involved in redox reactions in which the reaction species (oxidant) is reduced at the expense of the oxidation of the antioxidant (reductant). The FRAP assay measures the antioxidant effect of any substance in the reaction medium with reducing ability (19). Antioxidant potentials of various extracts obtained from roasted coffee were estimated by their ability to reduce TPTZ-Fe(III) complex to TPTZ-Fe(II) complex. In the present study, the trend for ferric ion reducing activities of extracts obtained from roasted coffee was shown in Fig. 1B. For roasted coffee, the absorbance clearly increased due to the formation of the Fe²⁺-TPTZ complex with increasing concentration. FRAP of green bean, roasted coffee and the positive control decreased in the following order: vitamin C (2.30) > medium (2.03) > green bean (1.90) > city (1.47) > french (1.16) at the concentration of 1,000 μg/mL. Similar to the results obtained from the ABTS radical scavenging activity assay, extracts of medium roasted coffee showed relatively strong ferric ion-reducing activity. A correlation between mean values of the total phenolics and antioxidant activities of medium roasted coffee deserves detailed attention, as phenolics in roasted coffee were likely capable of reducing ferric ions (Fig. 1B). Some researchers have reported similar correlations between phenolics and antioxidant activity measured by various methods (20). Our results showed that a progressive decrease in antioxidant activity and polyphenol content was observed with longer roasting time; therefore, the light roasted...
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Inhibitory effect of green and roasted coffees on lipid peroxidation

Recently, the study of lipid peroxidation is becoming increasingly of interest because the formation of cytotoxic products such as malondialdehyde (MDA) and 4-hydroxynonenal can influence cellular apoptosis and several human diseases (22). Therefore, in this assay, antioxidant activities of roasted coffee extracts on both ferric ion and vitamin C-induced lipid peroxidation in mouse whole brain homogenates were also confirmed. Results shown in Fig. 1C revealed that medium coffee extracts have a significant higher activity in suppressing lipid peroxidation in mouse whole brain homogenates. Although medium coffee extracts presented lower than catechin as a positive control at all concentrations, also noteworthy was that the inhibitory effect on lipid peroxidation was not induced by a single compound but just 70% ethanol extracts. Therefore, medium coffee might be a potential and natural antioxidant supplement for healthy foods and functional food products.

Neuronal cell protection of green and roasted coffees on H₂O₂-induced neurotoxicity

H₂O₂ has been reported to induce apoptosis in cells of the central nervous system (5,6). In this study, extracts obtained from roasted coffee were selected to investigate the neuroprotective effects against H₂O₂-induced damage because of strong antioxidant activity. Protective effects on H₂O₂-induced neuronal cell damage were examined by the MTT assay. H₂O₂ caused a decrease in cell viability (37%), but pretreatment of PC12 cells with increasing concentrations of the medium coffee extracts inhibited oxidative stress-induced cytotoxicity (Fig. 2A). Neuronal cell protective effect of medium coffee extracts at 62 μg/mL on oxidative injury was similar to that of 200 μM vitamin C. This study demonstrated that PC12 cell cytotoxicity through oxidative stress was suppressed by pretreatment with extracts obtained from medium roasted coffee. MTT dye reduction assay is based on the catalytic activity of some metabolic enzymes in intact mitochondria (23). Mitochondria may be one of the most sensitive primary targets of oxidative injury in neuronal cells (24). These results suggest that PC12 cell protection by medium roasted coffee may partially be due to mitochondrial protective mechanisms.
Protective effect of green and roasted coffees on H2O2-induced membrane damage

The neuronal membrane with polyunsaturated fatty acids is vulnerable to oxidative stress induced by ROS such as H2O2. Lipid peroxidation can alter the fluidity of the plasma membrane (18). LDH assay provided an estimate of the percentage of surviving PC12 cells. Green and roasted coffee extracts protected the integrity of the cellular membrane at all concentrations tested (Fig. 2B). Treatment with 200 μM H2O2 caused an increase in LDH released into the medium (65%). Compared with their effects in the corresponding H2O2 treatment group, green and all roasted coffee extracts dose-dependently decreased LDH released extracellularly. In addition, extracts of medium roasted coffee showed that the pattern of neuronal cell protection was similar to the MTT assay. The above results indicate that medium roasted coffee could protect PC12 cells against the membrane lesion induced by H2O2 (Fig. 2B). Therefore, these data suggest that PC12 cell protection by medium roasted coffee is partially due to the mitochondrial and cellular membrane protective mechanisms on H2O2-induced neurotoxicity.

Our results suggest that phenolics of roasted coffees might be inhibiting neuronal apoptosis, which is the ultimate consequence of these cellular dysfunctions. Especially, plentiful phenolics of medium-roasted coffees may also provide an added health benefit by reducing the risk of oxidative stress-induced neurodegenerative diseases.

Total phenolics and phenolic composition of green and roasted coffee beans

Phenolic compounds, such as flavonoids, phenolic acid and tannins, are considered to be major contributors to the antioxidant activity of natural plants. These antioxidants also possess diverse biological activities, including anti-inflammatory, anti-carcinogenic, and anti-neurodegenerative activities (25). Total phenolics of green and roasted coffee extracts were presented in Table 1. In addition, total phenolics of medium roasted coffee extracts were higher (9.77 mg GAE/g) than others. The extracts were subjected to further analysis by HPLC. The green and roasted coffee extracts contained various phenolic compounds. By comparing the retention time and UV spectra of these compounds with those of standards, 5-O-caffeoylquinic acid as the main phenolic was identified, followed by caffeic acid and syringic acid (Fig. 3). Furthermore, extracts from green coffee beans exhibited the highest 5-O-caffeoylquinic acid at 21.63 mg/100 g and in the following order medium (14.7 mg/100 g) > city (1.12 mg/100 g) > french (0.66 mg/100 g) (Table 1).
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In addition, when compared with other main phenolics as caffeic acid and syringic acid, the total sum of those in medium roasted coffee extracts was higher than another two coffee extracts. These results showed that many chemical component changes take place during roasting. Based on the results for the phenolic composition of extract from medium roasted coffee, we can conclude that these compounds (particularly 5-O-caffeoylquinic acid, caffeic acid and syringic acid) contribute to the protective effect on oxidative stress-induced neurotoxicity. The results obtained in this work are noteworthy, not only with respect to neuronal cell protections of roasted coffee extracts with phenolics but also with respect to compositional changes of phenolics due to roasting conditions.

Effect of 5-O-caffeoylquinic acid and epigallocatechin-3-gallate on lipid peroxidation

Today, green tea and coffee are the most widely consumed beverages by hundreds of millions of people around the world. Researches report that green tea has many health benefits because of physiological phenolics such as catechins. Catechins (flavan-3-ols) were the main phenolics in green tea, of which epigallocatechin-3-gallate (EGCG) represents approximately 59% of the total catechins (26). Therefore, we compared main phenolics of green tea and coffee on lipid peroxidation (Fig. 4). Compared with their effects on the corresponding H2O2 treatment group, EGCG and 5-O-caffeoylquinic acid dose-dependently decreased LDH released into medium (Fig. 4A). However, when compared with that of EGCG, membrane protective effect of 5-O-caffeoylquinic acid was lower. Similar to the results obtained from the LDH assay, EGCG and 5-O-caffeoylquinic acid both showed relatively strong inhibition of lipid peroxidation (Fig. 4B).
and the number of publications in this field increases every year. All of these publications include discussions of antioxidant activity capacity assays, comparisons of antioxidant activity of different phenolics and the chemical mechanism of scavenging ROS by phenolics (27,28). Because critical review papers on antioxidant activities of phenolics in green tea and coffee have been published, they are not a major point of discussion in this paper. Coffee and tea drinking are common worldwide, despite the internationally wide variety of drinking habits, e.g., type, frequency of intake, temperature, strength, etc. Although 5-O-caffeoylquinic acid showed lower antioxidant activity than that of EGCG at all concentration, this result was noteworthy because coffee has been one of the most popular beverages all over the world.

The physiological activities of roasted coffee extracts may be attributed to some phenolic compounds including 5-O-caffeoylquinic acid, caffeic acid and syringic acid. The content of 5-O-caffeoylquinic acid as a predominant phenolic compound was detected higher in medium roasted coffee extracts than others. Consequently, these data suggest that medium roasting condition in making tasty coffee from coffee beans may be more beneficial to human health, among them being the plentiful amounts of physiological phenolics including 5-O-caffeoylquinic acids.

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REFERENCES

