Quality Determination of Different Wholesale Cuts of Goat Carcass at Different Ages

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ABSTRACT - The experiment was conducted to find out the nutritive value and microbial status of different wholesale cuts of goat carcass. The meat sample (4 cuts from each animal × 3 different age groups × 3 animals in each group = 36 samples) was obtained from 1-, 2-, and 3-year aged goats. The whole sale cuts were shoulder, rack, loin and leg of each goat carcass. To assess the quality of meat sample, the general appearance, color, smell, juiciness, proximate composition, pH, total bacteria, coliform bacteria, and yeast were studied. The mean pH value of different cuts ranges from 5.65-5.69 didn’t differ significantly, but due to age differences the pH values (5.59-5.74) differed significantly (p < 0.01). The values of juiciness in different ages ranged from 32.24-42.10% which differed significantly (p < 0.01). The marbling of the cuts of rack portion was more pronounced than that of other cuts. The ranges of crude protein (CP) content of goat carcass (20.78-27.71%) differed significantly (p < 0.01) and leg portion contained higher CP than other portion. Fat contents of different cuts ranged from 2.66-11.47% differed significantly (P < 0.01). The moisture content of the carcass differed significantly which ranged from 69.20-73.31%. The ash content of the cuts of 1-year aged groups (0.99 ± 0.13%) was higher than that of other age groups and differed significantly (P < 0.01). The calcium (Ca) content did not differ significantly. The phosphorus (P) content was higher in one year old goat (0.15 ± 0.03%) than that of the goats of other ages. The total viable count (TVC) content of microorganisms ranging from 5.05-5.15 log cfu/g at different ages did not differ significantly. The coliform count (CC) of different cuts differed significantly (P < 0.01) which ranged from 2.56-3.05 log cfu/g; it also differed significantly (P < 0.05) in different ages (2.79-2.84 log cfu/g) and was higher in 1 year old goat carcass. The yeast count differed significantly in different cuts (P < 0.01) and ages (P < 0.05). From the study it is concluded that the age and different wholesale cuts have direct influence on quality of goat carcass.

Key words: Goat meat, Carcass quality, Wholesale cuts, Nutritive value, Microbial status

Chevon (goat meat) is one of the most widely eaten red meats in the world and enjoys great popularity in many developing countries, especially in Asia, Africa, and the Far East1. Goat meat is one of the most important protein sources for human consumption in Bangladesh. In almost all the communities of the country goat meat has long occupied a special place in the diet, for a variety of reasons including taste preference, prestige, religion, tradition and availability with the nutritional aspects being included more recently. Goat meat is a high-quality protein source. It is leaner than other red meats and its fat is less saturated, according to the USDA data2,3,4.

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There is a high demand of goat meat in the market due to its palatability, less fat content, tenderness and good flavor. All parts of the carcass are used for preparing a variety of rich dishes in many occasions. Goat meat consumption in the urban area has been increased to 23% between 1991/92 to 1995/965. Goat significantly contributes to the national GDP through the production of 1, 16000 MT of meat representing 25% of total red meat6. Goat meat is leaner than other red meats, as goats tend to deposit most of their body fat around omentum, mesentery and kidneys7. In general, goat carcass has more muscle and thicker muscle fiber bundles than sheep of similar age and weight group. The situation of meat supply in terms of handling, slaughtering and dressing of food animals in Bangladesh takes place in a very much disorganized way. Animals are slaughtered randomly and indiscriminately; the hygiene practice of ante mortem examination is not conducted9. Besides this, there is no any
system of grading in meat selling centers and slaughter houses on the basis of meat cuts and age differences in case of goat meat in Bangladesh. So, considering this point in mind this experiment was conducted to aware the consumers about the qualitative characters of different wholesale cuts of goat carcass and their differences in nutritive values among the cuts and also due to age differences. The present study was, therefore, set to get baseline information about the quality of goat meat. The objectives of this study were to determine the physical, chemical and microbial status of whole sale cuts of goat meat. The present study was, therefore, set to get baseline information about the quality of goat meat. The objectives of this study were to determine the physical, chemical and microbial status of whole sale cuts (shoulder, rack, loin and leg) of goat carcass, to compare the nutritional value among the four cuts, to find out the qualitative differences of goat carcass due to age differences.

Materials and Methods

Sample collection and preparation
Thirty six samples of goat meat of different wholesale cuts of different ages were studied in this experiment. Twelve kg of goat meat from freshly slaughtered goat were purchased from local market. Goat was slaughtered at 6 am in the municipal slaughter house of Mymensingh town. Fresh meat was collected and brought into the laboratory for processing. Fat, bone and fascia were removed from the cuts and fresh muscle was collected for the use of experiment. Then samples of meat (4 cuts from each animal × 3 different age groups × 3 animals in each group = 36 sample) were weighed in a balance and each of samples weighing 200 g meat.

pH Measurement
The samples (5 g each) were homogenized with 10 ml distilled water in a blender and the pH was measured using pH meter (Corning Model 250).

Determination of juiciness
The size of the block sample was usual size (2.5 × 2.5 cm) of meat which are generally consumed. About 100 g meat was prepared as block sample. The block meat sample (100 g) was cooked by heating only for 25 min in a saucepan. The cooked meat sample was put into three different plates, and weighed. Then the final weight was deducted from the initial weight.

Proximate Analysis
All proximate analyses were performed according to the AOAC procedures9. Moisture was determined by the oven drying method as outlined in procedure 24.0039. Protein was determined by the Kjeltech micro-Kjeldahl method, which was based on the Kjeldahl method as outlined in the AOAC procedure 2.0579. Fat was determined by soxhlet extraction using petroleum ether as outlined in procedure 7.0629. Ash was determined using procedure 31.0129. All determinations were done in triplicate then the mean value was reported.

Determination of Ca and P
For determination of Ca and P, 0.5 g of fresh meat samples were taken into a 100 ml digestion flask and then 10 ml of di-acid mixture (NHO₃: HClO₄ = 3:1) was added to the flask and then heated at 280°C for one hour. After cooling, distilled water was added to the digested sample and shake thoroughly. Then, the sample was filled up to 50 ml with distilled water and filtered through whatman (No. 1) filter paper.

For the calculation of calcium, 1 ml sample and 1 ml lanthanum chloride were mixed thoroughly, and the absorbance was measured by flame photometer. The calcium content of the sample was calculated from the standard curve. For the calculation of phosphorus, 5 ml of sample was mixed with 5 ml of ammonium molybdate and diluted with 2 ml of the stannous chloride (SnCl₂). The absorbance was measured by spectrophotometer 660 nm. The phosphorus values were calculated from the standard curve. The colorimetric method was employed to determine the P content of meat. Stannous chloride was used as a reducing agent and P content was measured at the wave length of 660 nm as outlined by Olsen and Sommers90.

Microbial Assessment
To observe the microbial status, Total Viable Count (TVC), Coliform Count (CC), Yeast count and Mould count were undertaken. The ground boneless lean goat meat samples were analyzed after keeping 2 months in 20°C. To determine these parameters, dilution blanks were prepared in the similar procedure which is described below:

Preparation of dilution blanks
One thousand ml of distilled water was taken into a sterilized flask. The phosphate buffer solution was prepared as per recommendation of the “Standard Methods for the Examination of Food Products”11. Then the buffer solution was added to the distilled water at the rate of 1.25 ml/1000 ml and the pH was adjusted to 7.2 ± 0.1. The dilution was then dispensed into several dilution bottles at the rate of 100 ml each and sterilized. Dilution blanks and the sample were shaken vigorously 25 times by up and down, 7 seconds for thorough mixing. Then by using a sterile pipette 1 ml portion was transferred to another 90 ml sterilized dilution blank. One and one-tenth ml portion of diluted sample from each dilution were placed into sterile petridishes aseptically in order to get the dilutions of 1:10, 1:10², 1:10³, 1:10⁴, 1:10⁵. Each of the dilutions was shaken as described earlier just before transferring from a dilution bottle into petridishes or another dilution bottle. The pipette was allowed for 2 to 3 seconds to drain off, then gently blown out the last drop
and touched the tip of the pipette to a dry spot on the glass. As soon as the dilutions were poured into petridishes, the mouth of the agar bottle was sterilized by flame and poured 10-15 ml of melted agar 40-45°C into each plate, slightly raising the lid of the petridishes. Agar was then mixed with the dilution by rotating and tilting the dish gently. The agar was then allowed to solidify.

**Total viable count (TVC)**

TVC was estimated by standard plate count technique. The plates were incubated at 37°C for 24 h and expressed as log cfu/g of sample. All plates for TVC and differential counts were prepared in duplicate.

After solidification of agar, the plates were inverted and placed in an incubator operated at 38°C for 48 hours. After incubation, the plates were taken out from the incubator and the plates which contained 30-300 colonies were selected for counting. Colonies were counted with the aid of a colony counter. The number of colonies was multiplied by the dilution and the total viable count per gram of sample was recorded.

**Determination of Coliform Count**

Coliforms were counted by using violet red bile agar (VRBA) plates. The plates were incubated at 44°C for 24 h. After incubation, the colonies were counted that having minimum diameter was about 0.5 mm. Finally, the total colony numbers of coliform were multiplied by the dilution and the counts per gram of sample were recorded.

**Determination of Yeast and Mould Count**

In this study Potato Dextrose Agar (PDA) was used to enumerate the yeast and mould count meat samples. After solidification of agar, the plates were inverted and incubated at 25°C for 3 days after incubation, the plates were taken out from the incubator and colonies were counted. Finally, the colony number of yeast and mould were multiplied by the dilution and the counts per gram of sample were recorded.

**Statistical analysis**

It was a 4 x 3 Factorial experiment and data were analyzed as per completely randomized design (CRD). In case of significant difference, least significant difference (LSD) was used to rank the treatment and/or interaction.

**Results and Discussion**

**Physico-chemical properties**

The physico-chemical attributes such as pH, juiciness, marbling, and nutritive value were observed and the results obtained are discussed below.

**pH**

The mean pH value of shoulder, Rack, Loin and Leg were 5.69, 5.66, 5.66 and 5.65 respectively (Table 1). There was no significant difference among the pH value of different cuts. The mean pH value of 1 year, 2 years and 3 years of chevon carcass were 5.59, 5.67 and 5.74 respectively (Table 2), there were significant (p < 0.01) difference on different ages. This result was similar to pH value of incised muscles was 5.65 in 6 hrs of slaughtering at ambient temperature. Gradual decline in pH was observed up to 48 hr of slaughtering²¹.

<table>
<thead>
<tr>
<th>Table 1. Effect of cuts on different parameters</th>
</tr>
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<tbody>
<tr>
<td>Parameters</td>
</tr>
<tr>
<td>pH</td>
</tr>
<tr>
<td>Juiciness</td>
</tr>
<tr>
<td>Moisture</td>
</tr>
<tr>
<td>Ash</td>
</tr>
<tr>
<td>Crude Protein</td>
</tr>
<tr>
<td>Ether extract</td>
</tr>
<tr>
<td>Ca²</td>
</tr>
<tr>
<td>P³</td>
</tr>
<tr>
<td>TVC (log cfu/g)</td>
</tr>
<tr>
<td>CC (log cfu/g)</td>
</tr>
<tr>
<td>Yeast (log cfu/g)</td>
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<tr>
<td>Mould</td>
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</tbody>
</table>

Means with different superscript in a row differ significantly
¹Calcium (Ca); ²Phosphorus (P); ³Total Viable Count (TVC); ⁴Coliform Count (CC); ⁵Least Significant Difference (LSD)
NS = Non-significant; **Significant at 1% level (P < 0.01)
Juiciness of meat is directly related to the intramuscular lipids and moisture content of the meat, but the water remaining in the cooked product is the major contributor to the sensation of juiciness during eating. The water holding capacity of the meat (WHC) influences juiciness of the meat. Meat with low WHC loses a lot of fluid in cooking and may taste dry and lack of succulence. The mean value of the juiciness of the meat (WHC) influences juiciness of the meat. Meat with low WHC loses a lot of fluid in cooking and may taste dry and lack of succulence. The mean value of the juiciness of the meat (WHC) influences juiciness of the meat. Meat with low WHC loses a lot of fluid in cooking and may taste dry and lack of succulence. The mean value of the juiciness of the meat (WHC) influences juiciness of the meat.

Table 2. Effect of age on different parameters

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Year</th>
<th>LSD value</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>P&lt;sup&gt;1&lt;/sup&gt;</td>
<td>1 Year (2 Teeth)</td>
<td>2 Years (4 Teeth)</td>
<td>3 Years (6 Teeth)</td>
</tr>
<tr>
<td>Juiciness</td>
<td>5.59&lt;sup&gt;a&lt;/sup&gt;</td>
<td>5.66&lt;sup&gt;b&lt;/sup&gt;</td>
<td>5.74&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Moisture</td>
<td>42.10&lt;sup&gt;a&lt;/sup&gt;</td>
<td>34.10&lt;sup&gt;b&lt;/sup&gt;</td>
<td>32.24&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Ash</td>
<td>73.31&lt;sup&gt;b&lt;/sup&gt;</td>
<td>69.21&lt;sup&gt;b&lt;/sup&gt;</td>
<td>69.65&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Crude Protein</td>
<td>0.99&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.92&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.91&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Ether extract</td>
<td>24.98&lt;sup&gt;a&lt;/sup&gt;</td>
<td>24.13&lt;sup&gt;b&lt;/sup&gt;</td>
<td>23.92&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Ca&lt;sup&gt;1&lt;/sup&gt;</td>
<td>7.47</td>
<td>7.81</td>
<td>8.22</td>
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<tr>
<td>P&lt;sup&gt;2&lt;/sup&gt;</td>
<td>0.04</td>
<td>0.04</td>
<td>0.02</td>
</tr>
<tr>
<td>TVC&lt;sup&gt;3&lt;/sup&gt;</td>
<td>0.15&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.12&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.14&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Yeast</td>
<td>5.05</td>
<td>5.05</td>
<td>5.15</td>
</tr>
<tr>
<td>Mould</td>
<td>2.84&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.80&lt;sup&gt;b&lt;/sup&gt;</td>
<td>2.79&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Moisture</td>
<td>1.35&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.40&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.48&lt;sup&gt;b&lt;/sup&gt;</td>
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<td>Mean value Significance</td>
<td></td>
<td>A×1</td>
<td>A×2</td>
</tr>
</tbody>
</table>

Means with different superscript in a row differ significantly

1<sup>Calcium (Ca)</sup>; 2<sup>Phosphorus (P)</sup>; 3<sup>Total Viable Count (TVC)</sup>; 4<sup>Coliform Count (CC)</sup>; 5<sup>Least Significant Difference (LSD)</sup>

NS = Non-significant; *Significant at 5% level (P < 0.05); **Significant at 1% level (P < 0.01)

Table 3. Combine effect of cuts and age on different parameter

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Year</th>
<th>Cuts and Interaction</th>
<th>LSD value</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>P&lt;sup&gt;1&lt;/sup&gt;</td>
<td>5.60</td>
<td>5.72</td>
<td>5.76</td>
<td>5.58</td>
</tr>
<tr>
<td>Juiciness</td>
<td>42.17</td>
<td>34.50</td>
<td>32.30</td>
<td>41.90</td>
</tr>
<tr>
<td>Moisture</td>
<td>72.43&lt;sup&gt;ab&lt;/sup&gt;</td>
<td>70.08&lt;sup&gt;bc&lt;/sup&gt;</td>
<td>67.85&lt;sup&gt;cd&lt;/sup&gt;</td>
<td>72.24&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Ash</td>
<td>0.81&lt;sup&gt;fg&lt;/sup&gt;</td>
<td>0.90&lt;sup&gt;de&lt;/sup&gt;</td>
<td>0.80&lt;sup&gt;e&lt;/sup&gt;</td>
<td>0.98&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td>CP&lt;sup&gt;4&lt;/sup&gt;</td>
<td>21.52</td>
<td>20.60</td>
<td>20.23</td>
<td>24.44</td>
</tr>
<tr>
<td>EE&lt;sup&gt;5&lt;/sup&gt;</td>
<td>6.99</td>
<td>7.15</td>
<td>7.40</td>
<td>11.07</td>
</tr>
<tr>
<td>Ca&lt;sup&gt;1&lt;/sup&gt;</td>
<td>0.04</td>
<td>0.07</td>
<td>0.01</td>
<td>0.04</td>
</tr>
<tr>
<td>P&lt;sup&gt;2&lt;/sup&gt;</td>
<td>0.15&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.14&lt;sup&gt;ab&lt;/sup&gt;</td>
<td>0.14&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.13&lt;sup&gt;ab&lt;/sup&gt;</td>
</tr>
<tr>
<td>TVC&lt;sup&gt;3&lt;/sup&gt;</td>
<td>5.13</td>
<td>5.17</td>
<td>5.08</td>
<td>5.01</td>
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<tr>
<td>Yeast</td>
<td>1.70&lt;sup&gt;cd&lt;/sup&gt;</td>
<td>2.81&lt;sup&gt;de&lt;/sup&gt;</td>
<td>2.60&lt;sup&gt;ef&lt;/sup&gt;</td>
<td>2.65&lt;sup&gt;ef&lt;/sup&gt;</td>
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<tr>
<td>Mould</td>
<td>1.30&lt;sup&gt;cd&lt;/sup&gt;</td>
<td>1.60&lt;sup&gt;de&lt;/sup&gt;</td>
<td>1.0&lt;sup&gt;d&lt;/sup&gt;</td>
<td>1.48&lt;sup&gt;de&lt;/sup&gt;</td>
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</tbody>
</table>

Means with different superscript in a row differ significantly

NS = Non-significant; *Significant at 5% level (P < 0.05); **Significant at 1% level (P < 0.01)

1<sup>Least Significant Difference (LSD)</sup>; 2<sup>Crude Protein (CP)</sup>; 3<sup>Ether Extract (EE)</sup>; 4<sup>Calcium (Ca)</sup>; 5<sup>Phosphorus (P)</sup>; 6<sup>Total Viable Count (TVC)</sup>; 7<sup>Coliform Count (CC)</sup>

Juiciness of meat is directly related to the intramuscular lipids and moisture content of the meat, but the water remaining in the cooked product is the major contributor to the sensation of juiciness during eating. The water holding capacity of the meat (WHC) influences juiciness of the meat. Meat with low WHC loses a lot of fluid in cooking and may taste dry and lack of succulence. The mean value of the juiciness in shoulder, rack, loin, and leg were 36.36, 36.00, 36.03 and 36.21, respectively (Table 1) which did not differ significantly. Higher intramuscular fat contents are associated with increased tenderness in flesh. It also affects juiciness in meat, as flavor factors, stimulates salivary flow is concentrated in fatty tissue. The mean value of juiciness in 1 year, 2 years
and 3 years were 42.10, 34.10 and 32.24, respectively (Table 2), which differ significantly (p < 0.01).

**Marbling**
Marbling of different cuts such as shoulder, rack, loin and leg were observed by eye estimation. The rack portion of goat carcass had more marbling than others, but on an average the muscles of all cuts had less marbling which is desirable. The marbling was slightly different among different ages of carcasses such as 1 year, 2 years and 3 years. Marbling is higher with the increase of age, but not significantly higher.

**Proximate Composition**
Shoulder, rack, loin and leg from fresh boneless lean chevon were analyzed for the determination of chemical composition. Crude protein, ether extract, moisture, ash, calcium and phosphorus contents were obtained from the analyses. The results are discussed below.

**Moisture**
The moisture tends to be less as the animal becomes more fat. Slaughter age had a significant effect on moisture, protein, iron, calcium and pH. Fat, protein, and iron contents increased with advancing slaughter age, while moisture content decreased. The mean value of moisture contents of shoulder, rack, loin and leg were 70.12%, 69.60%, 70.06% and 73.12%, respectively, which are shown in Table 1. The cuts of leg (73.12%) significantly (p < 0.01) differ from other cuts of the carcass. The mean values of moisture contents of shoulder, rack, loin and leg were decreased. The mean value of moisture of 1 year, 2 years and 3 years were 73.31%, 69.21% and 69.65%, respectively (Table 2), where cuts of one year (73.31%) age significantly differ (p < 0.01) from the cuts of other ages. The moisture content for donkeys was lower (68.35-74.72%), significantly (p < 0.01) from the cuts of other ages. The mean value of moisture of 1 year, 2 years and 3 years of goat carcass were 24.98, 24.13 and 23.92%, respectively. The cuts of one year age significantly differed (p < 0.05) from the carcass of other ages (Table 2). The protein content was high for donkey meat, than the CP in beef. In beef, protein content was of 20% and 1.09% for donkeys, respectively. The cuts did not have any significant difference.

**Fat**
Fat deposition in the carcass or in the abdominal cavities was increased by the influence of live weight at slaughter. Fat content of the cut are shown in Tables 1 and 2. The fat content of shoulder, rack, loin, and leg were 0.04, 0.03, 0.03 and 0.04%, respectively (Table 2) which were also differ significantly (p < 0.01). The fat content of rack (11.47%) was the highest among the different ages of carcass. Chevon is also attractive to health conscious consumers due to its low fat compared to other traditional red meats such as lamb, beef, and pork.

**Calcium**
Calcium content is lower in lean flesh of chevon (0.01-0.02%). The mean calcium contents of shoulders, rack, loin, and leg were 0.04, 0.03, 0.03 and 0.04%, respectively (Table 1). The cuts did not have any significant difference. The mean value of different ages are shown in Table 2, they also did not have any significant difference.

**Ash**
The ash values of cuts are presented in Tables 1 and 2. The mean ash value of shoulder, rack, loin, and leg was 0.84%, 0.98%, 0.92% and 1.03%, respectively (Table 1), which were also differ significantly (p < 0.01). The ash content of 1 year, 2 years and 3 years of age was 0.99%, 0.92% and 0.91%, respectively (Table 2) which were also differ significantly. The ash content of 1 year age, 0.99% was higher than that of the cuts of other ages. The total ash content for donkey cuts was the highest (5.10-8.19%) when compared to that of beef, reported 1.5%23, while other findings were slightly lower, reporting a mean value of 1.09%27. Ash (inorganic minerals) content of muscle tissue is approximately 1%. Ash content accurately reflects the mineral content, but does not differentiate among minerals. Because of the relatively low content of minerals in fatty tissues, the fat level also indirectly influences the mineral or ash content of meat and meat products24.
Phosphorus

Meat is a good source of phosphorus compared to calcium\(^2\). The mean phosphorus contents of shoulder, rack, loin, and leg were 0.14, 0.13, 0.14 and 0.12\%, respectively which did not differ significantly. The mean value of phosphorus of 1 year, 2 years, and 3 years were 0.15, 0.12 and 0.14\%, respectively (Table 2) which differ significantly (p < 0.01). The mean phosphorus content of the cuts of 1 year age (0.15\%) was higher than the cuts of other ages.

Microbial Status

Total Viable Count (TVC) of microbes

The average total viable count (TVC) per gram of meat was recorded as 1.32 × 10\(^7\) cfu/g\(^3\). The mean values of the Total Viable Count (TVC) per gram of shoulder, rack, loin, and leg were 5.13, 5.12, 5.02 and 5.08 log cfu/g, respectively (Table 1). There were significant differences among the cuts that the general viable count of fresh meat tissue at 35°C should be less than 10\(^7\)cfu/g\(^3\). The mean value of TVC per gram of goat meat sample of 1 year, 2 years and 3 years of carcasses were found 5.05, 5.05, 5.15 log cfu/g, respectively presented in Table 2. There were no significant differences of TVC in the carcasses of different ages.

Coliform Count

The mean values of coliform count in shoulder, rack, loin, and leg were 2.56, 2.17, 2.78 and 3.05 log cfu/g, respectively (Table 1). The result differed significantly (p < 0.01). The average count of total organism on the internal surfaces was log 7.21 and the external surface was log 5.53 per square cm, respectively\(^6\). The mean value of coliform count in different ages such as 1 year, 2 years and 3 years were 2.84, 2.80 and 2.79 log cfu/g, respectively (Table 2) which have significant difference (p < 0.05). The cuts of 1 year age contained higher coliform count than others.

Yeast and Mould count

The mean values of yeast and mould count of shoulder, rack, loin, and leg are shown in Table 1. There were a significant differences (p < 0.05) in yeast content in different cuts. No mould was found in the samples. The mean value of yeast and mould count of 1 year, 2 years and 3 years of carcasses are shown in Table 2. The mean yeast number in 1 year, 2 years and 3 years were 1.35, 1.40 and 1.48 log cfu/g, respectively, which showed significant difference (p < 0.05) but no mould was found in carcass of different ages.

Conclusion

From the present study it can be concluded that the age of goat has the influence on nutrient content of goat carcass. Juiciness, moisture, crude protein, and ash value was found significantly higher (p < 0.05; p < 0.01) in 1 year aged goat meat compared to 2 and 3 years aged goat meat. It may further be concluded that there was also little change in nutrient content of different wholesale cuts. Moisture, ash and crude protein content of leg cut were significantly higher (p < 0.01) than that of other cuts. The total viable count of microorganisms ranging from 5.05-5.15 log cfu/g at different ages did not differ significantly (p > 0.05). Microbial quality of meat depends upon the slaughter place, handling and storage condition. Good hygienic management leads to good microbial quality of meat. According to the findings of this study consumer can prefer 1 year aged goat meat and also meat from leg cut for better nutritional quality. However, it needs further investigation/study.
Reference